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# Assessment of quinolone resistance in bacteria isolated from corneal ulcers in dogs

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# Abstract

Corneal ulcers or ulcerative keratitis is the loss of continuity of corneal epithelium leading to exposure of corneal stroma. It is a highly painful condition in dogs. The common aetiological agents responsible for this condition are bacteria and for treatment, antibiotics are widely used. Indiscriminate and prolonged use of antibiotics can cause antimicrobial resistance (AMR) in bacteria. The present communication deals with the isolation of bacteria from corneal ulcers in dogs and the assessment of quinolone resistance. A total of 15 corneal swab samples were collected from dogs suffering from corneal ulcers, presented to the Department of Veterinary Surgery and Radiology, Teaching Veterinary Clinical Complex, College of Veterinary and Animal Sciences, Mannuthy. They were inoculated on to brain heart infusion agar as well as blood agar and incubated at 37 °C for 24 h for the isolation of aerobic bacteria. Eighteen isolates obtained were then subjected to antibiotic susceptibility test by disc diffusion method using the quinolone group of antibiotics, viz. ciprofloxacin, enrofloxacin, ofloxacin and moxifloxacin. Majority of the isolates exhibited varying resistance to these antibiotics. Polymerase chain reaction was performed to amplify gyrA gene, which is reported to be one of the reasons responsible for quinolone resistance with respect to its mutation. Seven out of the 18 isolates amplified gyrA gene. The amplicon obtained corresponding to a representative sample, which was found resistant by disc diffusion method was sequenced and a single point mutation was detected.

Keywords: Quinolones, antimicrobial resistance, bacterial isolates, corneal ulcers

Antimicrobial resistance (AMR) is an emerging public health concern, which is mainly due to the indiscriminate use of antibiotics over a long period of time. Since dogs are the best companion animal, they are considered to be potential carriers of microbes with AMR genes to human beings

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(Guardabassi *et al.*, 2004). Corneal ulcers or ulcerative keratitis is one of the most common and painful eye affections in dogs. It is defined as the loss of continuity of corneal epithelium leading to exposure of corneal stroma. The predominant bacteria associated with corneal ulcers are Gram positive, *viz. Staphylococcus* spp. (Murali *et al.*, 2017), *Streptococcus* spp., *Corynebacterium* spp. and *Bacillus* spp. while, the Gram negative organisms included *Pseudomonas* spp., *Klebsiella* spp., *E. coli* and *Neisseria* spp. (Moore and Naisse, 1999). Hence, isolation of bacteria from corneal ulcers in dogs and determination of antibiotic susceptibility pattern is of great importance.

Corneal swab samples collected were inoculated onto brain heart infusion agar and blood agar and incubated at 37 °C for 24 h. Eighteen isolates could be obtained with twelve as pure cultures and three as mixed, each with two types of colonies. On Gram's staining, seven were Gram positive cocci, five Gram positive bacilli, five Gram negative bacilli and one was identified as Gram negative diplococci. Based on colony morphology, Gram's staining and biochemical tests described by Barrow and Feltham (1993), the isolates were identified as *Staphylococcus* spp. (five), *Corynebacterium* spp. (five), *Streptococcus* spp. (two), *Pseudomonas* spp. (two), *Klebsiella* spp. (two), *Neisseria* spp. (one) and *E. coli* (one). Similar results were recorded by Hewitt *et al.* (2020).

Antibiotic susceptibility testing of the isolates was carried out by disc diffusion method in Mueller-Hinton agar, using the quinolone group of antibiotics, *viz*. ciprofloxacin, moxifloxacin, enrofloxacin and ofloxacin. Antimicrobial susceptibilities for all isolates were determined according to Clinical and Laboratory Standards Institute guidelines

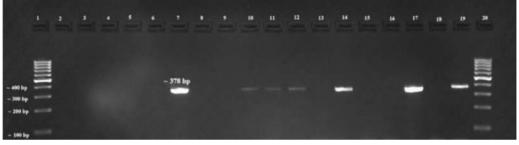


Fig. 1. Agarose gel electrophoresis depicting amplicons specific to *gyrA* gene Lane 1, 20 – 100 bp ladder Lane 2 to 19 – samples (S1-S6; S7C, S7N, S8-S11; S12C, S12S, S13, S14, S15C, S15S)

Query	109	GCACGTGACGGCCTGAAGCCGGTGCACCGCCGTGTGCTTTATGCCATGAGCGAGC	168
Sbjct	1	GCACGTGACGGCCTGAAGCCGGTGCACCGCCGTGTGCTTTATGCCATGAGCGAGC	60
Query	169	AACGACTGGAACAAGCCCTACAAGAAATCCGCCCGTGTGGTCGGCGACGTGATCGGTAAG	228
Sbjct	61	AACGACTGGAACAAGCCCTACAAGAAATCCGCCCGTGTGGTCGGCGACGTGATCGGTAAG	120
Query	229	TACCACCCGCACGGCGACACCGCGGTCTACGACACCATCGTGCGCATGGCGCAGCCGTTC	288
Sbjct	121	TACCACCGCACGGCGACACCGCGGTCTACGACACCATCGTGCGCATGGCGCAGCCGTTC	180
Query	289	TCGCTGCGCTACATGCTGGTAGACGGCCAGGGCAACTTCGGTTCGGTGGACGGCGACAAC	348
Sbjct	181	TCGCTGCGCTACATGCTGGTAGACGGCCAGGGCAACTTCGGTTCGGTGGACGGCGACAAC	240
Query	349	GCCGCAGCCATGCGATACACCGAAGTGCGCATGGCCAAGCTGGCCCACGAACTGCTGGCG	408
Sbjct	241	GCCGCAGCCATGCGATACACCGAAGTGCGCCATGGCCCAAGCTGGCCCATGAACTGCTGGCG	300
Query	409	GACCTGGAAAAGGAAACCG 427	
Sbjct	301	GACCTGGAAAAGGAAACCG 319	

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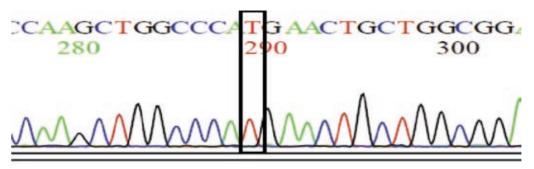


Fig. 3 Chromatogram of the sequenced isolate

(CLSI, 2020). It revealed that majority of them (88.89 per cent) were resistant to the quinolone group of antibiotics, *viz*. ciprofloxacin, moxifloxacin, enrofloxacin and ofloxacin. Only one isolate each of *Corynebacterium* spp. and *Staphylococcus* spp. were found to be sensitive to quinolones. Hindley *et al.* (2015) reported a similar finding in which isolates from canine ulcerative keratitis were found to be resistant to quinolone group of antibiotics.

Genomic DNA was extracted from these cultures and polymerase chain reaction (PCR) was performed to amplify the antibiotic resistance gene, gvrA in these isolates. The primersequencesusedfortheamplificationofthe genewereas follows (5'-3'): gyrA Forward primer, AGTCCTATCTCGACTACGCGAT;gyrA Reverse primer, AGTCGACGGTTTCCTTTTCCAG (Lin et al., 2012). After performing the PCR, products were electrophoresed in 2 per cent agarose gel by submarine agarose gel electrophoresis and visualized by gel documentation system (Bio-Rad, USA). Only seven isolates were found to amplifythisgene(Fig.1), which showed amplicon of size 378 bp, while the rest of the isolates failed to amplify this gene. Osman et al. (2016) and Rasool et al. (2020) also reported similar finding in which all the isolates in their study did not amplify the gyrA gene. Since the mutation in gyrA is the commonly reported reason for quinolone resistance, gyrA specific amplicon of representative isolate of Pseudomonas spp. in the present study, was selected for sequencing. The sequence was analysed with that of the reference strain, Pseudomonas aeruginosa PAO1 (Nouri et al., 2016). A single point mutation in nucleotide sequence was detected in the sample sequenced (Fig. 2 and 3) but it did not alter the amino acid pattern. Hence,

the mutation detected was silent. The mutation reported by Lin *et al.* (2012) was substitution of Thr at position 83 with lle in *gyrA*. However, the association of mutation recorded in present study with AMR need further investigation. A comprehensive study is required to unravel the effect of mutations in specific gene or presence of other AMR genes to understand the resistance mechanism against quinolones.

## Summary

The present study revealed that the most commonly isolated bacteria from ulcerative keratitis in dogs were Gram positive with *Staphylococcus* spp. and 88.89 per cent of the isolates were found to be resistant to quinolone group of antibiotics. The amplicon of representative isolate sequenced, possessed a silent mutation. Hence, further studies are warranted for better understanding of the quinolone resistance mechanism.

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#### **Conflict of interest**

The authors declare that they have no conflict of interest.

## References

# Barrow, G.I. and Feltham, R.K.A. 1993. *Cowan* and Steel's Manual for The Identification of Medical Bacteria. (3<sup>rd</sup> Ed.). Cambridge University Press, UK, 331p.

- CLSI [Clinical and Laboratory Standards Institute]. 2020. Performance Standards for Antimicrobial Disk and Dilution Susceptibility Tests for Bacteria Isolated from Animals: (5<sup>th</sup> Ed.). CLSI Supplement VET01S; CLSI: Wayne, PA, USA. 216p.
- Guardabassi, L., Schwarz, S. and Lloyd, D. H. 2004. Pet animals as reservoirs of antimicrobial-resistant bacteria. *J. Antimicrob. Chemother.* **54**(2): 321-332.
- Hewitt, J. S., Allbaugh, R. A., Kenne, D. E. and Sebbag, L. 2020. Prevalence and antibiotic susceptibility of bacterial isolates from dogs with ulcerative keratitis in midwestern United States. *Front. Vet. Sci.* [Online]. 7: 583965. Available: http:// doi.org/10.3389/fvets.2020.583965 [20 Nov. 2020].
- Hindley, K. E., Groth, A. D., King, M., Graham, K. and Billson, M. 2015. Bacterial isolates, antimicrobial susceptibility and clinical characteristics of bacterial keratitis in dogs presenting to referral practise in Australia. *J. Vet. Ophthalmol.* **19** (5): 418-426.
- Lin, D., Foley, S. L., Qi, Y., Han, J., Ji, C., Li, R., Wu, C., Shen, J. and Wang, Y. 2012. Characterization of antimicrobial resistance of *Pseudomonas aeruginosa* isolated from canine infections. *J. Appl. Microbiol.* **113** (1): 16-23.

- Moore, C.P. and Nasisse, M.P. 1999. The canine glaucoma. In: Gelatt, K.N. (ed.), *Veterinary Ophthalmology.* (3<sup>rd</sup> Ed.). Lippincott Williams and Wilkins, Philadelphia, pp. 259-290.
- Murali, A. V., Mani, B. K., Mini, M., Joseph, S. and Unny, N.M. 2017. Characterisation of *Staphylococcus aureus* isolated from dogs with corneal diseases. *J. Vet. Anim. Sci.* **48**(2): 16-19.
- Nouri, R., Ahangarzadeh Rezaee, M., Hasani, A., Aghazadeh, M. and Asgharzadeh, M. 2016. The role of *gyrA* and *parC* mutations in fluoroquinolones-resistant *Pseudomonas aeruginosa* isolates from Iran. *Braz. J. Microbiol.* **47**: 925-930.
- Osman, K., Badr, J., Al-Maary, K.S, Moussa, I.M.I., Hessain, A.M., Girah, Z.M.S.A., Abo-shama, U.H., Orabi, A. and Saad, A. 2016. Prevalence of the antibiotic resistance genes in coagulase positive negative Staphylococcus and in chicken meat retailed to consumers. Front. Microbiol. [Online]. 7:1846. https://doi.org/10.3389/ Available: fmicb.2016.01846 [22 Nov. 2016].
- Rasool, K. H, Hussein, N. H. and Taha, B. M. 2020. Molecular detection of *gyrA* gene in *Salmonella enterica* serovar Typhi isolated from typhoid patients in Baghdad. *Pak. J. Biol. Sci.* **23**: 1303-1309.