



# Molecular characterisation of virulence genes in *Staphylococcus aureus* associated with clinical bovine mastitis

 Anugraha Mercy Easaw<sup>1\*</sup>,  K. Vijayakumar<sup>2</sup>,  K. Justin Davis<sup>3</sup>,  V.H. Shyma<sup>3</sup> and S. Surya<sup>4</sup>

Department of Veterinary Epidemiology and Preventive Medicine  
College of Veterinary and Animal Sciences, Mannuthy, Thrissur- 680 651  
Kerala Veterinary and Animal Sciences University, Kerala, India.

Citation: Easaw, A.M., Vijayakumar, K., Davis, J., Shyma, V.H. and Surya, S.2022. Molecular characterisation of virulence genes in *Staphylococcus aureus* associated with clinical bovine mastitis. *J. Vet. Anim. Sci.* **53**(2): 279-284

DOI: <https://doi.org/10.51966/jvas.2022.53.2.279-284>

Received: 16.11.2021

Accepted: 14.12.2021

Published: 30.06.2022

## Abstract

*Staphylococcus aureus* is the most frequently isolated pathogen from bovine mastitis including subclinical, clinical, and chronic infections. Virulence factors possessed by *S. aureus* aid in causing infection and inflammation by producing toxins and proteins, which are responsible for the pathogenesis of the disease. In the current study out of 51 animals presented with clinical mastitis, *S. aureus* was isolated from the milk of 18 animals. *S. aureus* was confirmed by genus and species level identification using polymerase chain reaction. Molecular characterization of selected virulence genes including thermonuclease (*nuc*) and Panton Valentine Leucocidin (PVL) was performed in all the *S. aureus* isolates. Presence of *nuc* gene was observed in all the isolates (100 %) of *S. aureus*. No isolates were found to be positive for the presence of PVL gene. Profiling the virulence genes is an important tool for epidemiological studies of mastitis, which can be employed for the prevention and control of the disease.

**Keywords:** Mastitis, *Staphylococcus aureus*, virulence genes

Mastitis can be defined as a complex disease which involves interaction between the host anatomy and physiology, different causative pathogens and environmental factors such as animal husbandry, hygiene and sanitation. In dairy industry worldwide, *S. aureus* is said to be the most common causative organism responsible for bovine mastitis (Miles *et al.*, 1992). The infected quarter of affected cows is considered to be the main reservoir for *S. aureus* infection in the herd. The different factors like evasion of immune mechanism of host, invasion and infection of host tissue, spread of bacteria and acquisition of the required nutrients by the pathogenic organism like *S. aureus* could be attributed to its virulence factors (Haveri *et al.*, 2005). The *nuc* gene is

1. M.V. Sc Scholar
2. Professor and Head,
3. Assistant Professors, Department of Veterinary Epidemiology and Preventive Medicine
4. Assistant Professor, Department of Veterinary Microbiology College of Veterinary and Animal Sciences, Mannuthy, Thrissur- 680651

Corresponding author Email id: [mercyvet2013@gmail.com](mailto:mercyvet2013@gmail.com) , Ph no: +916238535639

Copyright: © 2022 Easaw *et al.* This is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

said to be ubiquitous in all organisms belonging to genus *Staphylococcus*. It is said that regardless of the thermonuclease activity all *Staphylococcus* spp., except *S. scuri*, possess *nuc* gene (Sasaki *et al.*, 2007). Panton Valentine Leucocidin is a cytotoxin, which causes disruption of cell membranes creating pores on polymorphonuclear neutrophils (Kaneko and Kameo 2004). Panton Valentine Leucocidin positive *S. aureus* was found to be responsible for causing mastitis in many countries and it was observed that even though PVL acts strongly on human polymorphonuclear cells, weak activity was observed on bovine neutrophils too (Peton and Le Loir, 2014).

The study of the important virulence factors aids in the identification of their role in bovine mastitis. Each specific virulence factors have role in each stage and type of infection, and it is observed that not all strains of *S. aureus* possess the same virulence factors. Hence, study of virulence factors in *S. aureus* is useful in molecular epidemiological studies of bovine clonal types for the effective prevention and control of mastitis (Fitzgerald *et al.*, 2000). Hence, this study was envisaged for the molecular detection of virulence genes, *nuc* and *PVL* from clinical mastitis caused by *S. aureus*.

## Materials and methods

Milk samples were collected from 51 animals affected with clinical mastitis and subjected to identification of organisms by morphological characterization, colony characteristics on selective media and biochemical reactions (Barrow and Feltham, 1993; Quinn *et al.*, 2013). Isolates presumptive of *S. aureus* were further subjected for confirmation by molecular characterisation. The DNA extraction was performed by snap chill method or heat lysis method as described by Vijayakumar and Jose (2021). All the isolates of *S. aureus* obtained in the study were subjected to genotypic characterisation by amplification of 16S rRNA and 23S rRNA genes for molecular confirmation of *Staphylococcus* spp. and *S. aureus* respectively. The presence of selected virulent factor genes *viz.* *nuc* for thermonuclease and *PVL* for Panton Valentine Leucocidin were determined by polymerase

chain reaction (PCR) using specific primers as shown in Table 1. The reagents and chemicals used for the PCR were Emerald Amp Fast PCR master mix (2X PCR Smart mix, Takara, Japan) forward and reverse primer set (100nM/ml, Sigma Aldrich) and sterile nuclease free water. All the primers were reconstituted in sterile nuclease free water to a final concentration of 10 pmol/μl and stored at -20 °C.

The PCR were performed using the programmable S1000 Thermal cycler, BioRad, USA. Polymerase chain reaction was performed in a total volume of 25μl reaction mixture by combining the reagents. The PCR conditions were optimized by setting different time temperature combinations for annealing processes (Tables 2, 3). The combination that gave the best result for amplification was selected for carrying out further PCR.

After completion of PCR, amplified products were subjected to submarine agarose gel electrophoresis.

## Results and discussion

In the present study, out of the samples from 51 animals affected with clinical mastitis, 35 samples yielded growth and a total 38 bacterial isolates were obtained wherein, three samples yielded two different types of growth. Out of the 38 organisms isolated during this study, 27 were contagious pathogens (71.05 %) which included 18 isolates of *S. aureus* (47.37 %), 6 isolates of coagulase negative staphylococci (15.79 %) and 3 isolates of *Micrococcus* spp. (7.89 %). The environmental pathogens (28.95 %) that were isolated included 11 coliforms, out of which 6 isolates were *E. coli* (15.79 %) and 5 isolates were *Klebsiella* spp. (13.16 %). *Staphylococcus aureus* was isolated as the major pathogen from bovine mastitis cases and the result was in accordance with the findings by Verma *et al.* (2018), and Workineh *et al.* (2002) who reported the prevalence of *S. aureus* as 42.55 per cent and 40.5 per cent, respectively. Studies conducted on bovine clinical mastitis by Rathish *et al.* (2015) in Thrissur district also revealed *S. aureus* to be the most common pathogen isolated. The present study did not concur with the findings of Fadlilmula *et al.* (2009), who found lower prevalence of *S. aureus*

**Table 1.** Details of primers used in PCR

Organism / Virulence genes	Genes	Primer sequence	Amplicon size (bp)	Reference
<i>Staphylococcus</i> spp.	16S rRNA	F: AACTCTGTTATTAGGGAAGAA CA R: CCACCTTCCTCCGGTTTGTCCACC	756	Ciftci <i>et al.</i> (2009)
<i>Staphylococcus aureus</i>	23S rRNA	F: GGA CGA CAT TAG ACG AAT CA R: CGG GCA CCT ATT TTC TAT CT	1318	El - Razik <i>et al.</i> (2010)
Thermonuclease	nuc	F: GCCAAGCCTTGACGAACATAAGC R: GCGATTGATGGTGATACGGTT	279	Brakstad <i>et al.</i> (1992)
Panton Valentine Leucocidin	PVL	F: GCTGGACAAAACCTTCTTGAATAT R: GATAGGACACCAATAAATTCTGGATTG	85	Pajic <i>et al.</i> (2014)

**Table 2.** PCR protocol for the amplification for characterization of *S. aureus*

Sl. No	PCR Programme		Temperature – Time Protocol	
			16S rRNA	23S rRNA
1.	Initial Denaturation		94 °C for 5 min	94 °C for 5 min
2.	Denaturation	30 cycles	94 °C for 45 sec	94 °C for 45 sec
3.	Annealing		56.9 °C for 45 sec	55.8 °C for 45 sec
4.	Extension		72 °C for 90 sec	72 °C for 90 sec
5.	Final extension		72 °C for 10 min	72 °C for 10 min
6.	Hold		4 °C Until use	4 °C Until use

**Table 3.** PCR protocol for the amplification of virulence genes of *S. aureus*

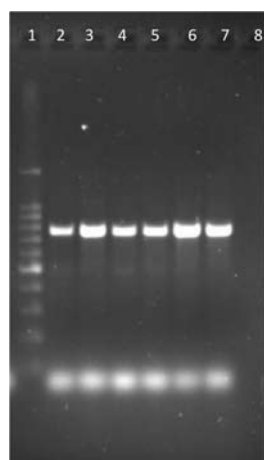
Sl. No	PCR Programme		Temperature - Time Protocol	
			nuc	PVL
1.	Initial Denaturation		94 °C for 5 min	95 °C for 5 min
2.	Denaturation	30 cycles	94 °Cfor 1 min	95 °C for 30 sec
3.	Annealing		55 °C for 30 sec	59 °C for 30 sec
4.	Extension		72 °C for 90 sec	72 °C for 60 sec
5.	Final extension		72 °C for 3.5 min	72 °C for 5 min
6.	Hold		4 °C Until use	4 °C Until use

of 9.8 per cent in their study. Jose *et al.* (2021) reported in their studies a lower prevalence of coliforms, *Klebsiella* spp. (7.40 per cent) and *E. coli* (4.47 per cent), isolated from clinical bovine mastitis which was consistent with our study.

Hence, from clinical bovine mastitis cases, 18 isolates of *S. aureus* were identified by biochemical methods and subjected to molecular confirmation by polymerase chain reaction after DNA extraction by snap chill method. This method was used for extraction of bacterial DNA from cases of bovine mastitis by many researchers (Shah *et al.*, 2020; Nazir *et al.*, 2017). Lange *et al.* (2015) performed species level identification of *Staphylococci* and concluded that 16S rRNA sequencing is an accurate method for the identification of

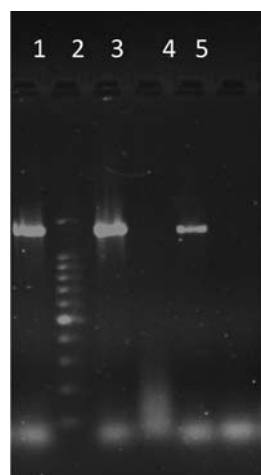
staphylococci from bovine mastitis. But due to the lack of heterogenicity in 16S rRNA gene, it would be difficult to identify and discriminate different *Staphylococcus* species (Petti *et al.*, 2008).

Stephan *et al.* (2001) used 23S rRNA for the species level identification of *S. aureus* from bovine mastitis, and all the 34 isolates could be identified as *S. aureus*. Hence, all the bacterial isolates presumed to be *Staphylococcus* spp. were subjected to PCR by targeting 16S rRNA and for the species level identification of the isolates, 23S rRNA was targeted. From clinical bovine mastitis, 18 isolates were identified and confirmed as *S. aureus* respectively (Fig. 1 and Fig. 2).



**Fig. 1. Agarose gel electrophoresis of 16S rRNA specific PCR of *Staphylococci* spp.**

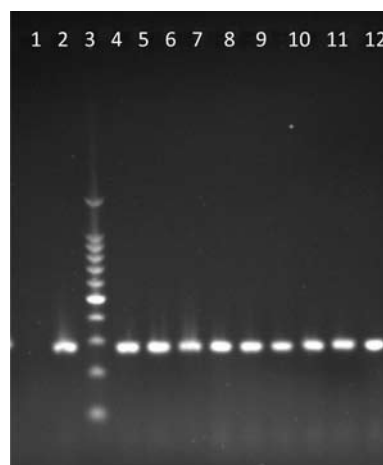
Lane 1 – Ladder  
Lane 7 – Positive control  
Lane 8 – Negative control  
Lane 2,3,4,5,6,7 – Positive samples (756 bp)



**Fig. 2. Agarose gel electrophoresis of 23S rRNA specific PCR of *S. aureus***

Lane 1,3 – Positive samples (1318 bp)  
Lane 2 – Ladder  
Lane 4 – Negative control  
Lane 5 – Positive control

The virulence factors thermonuclease and Pantone-Valentine Leukocidin coded by *nuc* and *PVL* respectively were screened. The *nuc* gene is commonly employed for the species level identification of *S. aureus*. The positive amplicon size of 279 bp for *nuc* gene was detected in all the 18 isolates (100 per cent) of *S. aureus* (Fig.3). This was in accordance with many studies wherein, *nuc* gene was used for



**Fig. 3. Agarose electrophoresis of *nuc* gene specific PCR of *S. aureus***

Lane 1 – Negative control  
Lane 2 – Positive control  
Lane 3 – Ladder  
Lane 4,5,6,7,8,9,10,11,12 – Positive samples (279 bp)

the identification of *S. aureus* organism isolated from bovine mastitis (Ciftci *et al.*, 2009). *S. aureus* possess *nuc* gene which has species specific sequences which on amplification have the potential for detection as well as identification of *S. aureus* from infections (Brakstad *et al.*, 1992; Costa *et al.*, 2004).

Panton Valentine Leucocidin is a phage encoded virulence factor which has major public health significance. Screening was performed for the presence of *PVL* in 18 isolates of *S. aureus* from bovine mastitis and none of them was found to have the presence of the gene. Similarly, absence of *PVL* was observed in other studies (Patel *et al.*, 2021; Prashanth *et al.*, 2011). Varying occurrence of *PVL* in bovine mastitis is observed throughout India, as high as 41.6 per cent (Mitra *et al.*, 2013) and as low as 10.53 per cent (Shrivastava *et al.*, 2018). Identification of *PVL* in bovine isolates is a rare finding and the frequency observed in other studies on bovine mastitis was attributed to the contamination of milk by milkmen carrying *PVL* containing *S. aureus* isolates (Fluit, 2011; Shrivastava *et al.*, 2018). Unlike other leukotoxins, *PVL* is found to have weak action on bovine neutrophils, and this could be the probable reason for the absence of *PVL* in bovine strains and for its presence in human strains of *S. aureus* (Prevost *et al.*, 1995).

## Conclusion

The present study concluded that there is presence of virulence gene in *S. aureus* associated with bovine mastitis isolated from Thrissur district. Further study of different virulence genes in a large study population must be conducted for extrapolating the data, to use it for epidemiological investigation and for the prevention and control of mastitis caused by *S. aureus*.

## Conflict of interest

The authors declare that they have no conflict of interest.

## References

- Barrow, C.I. and Feltham, R.K.A. 1993. Cowan and Steel's Manual for the Identification of Medical Bacteria. (3<sup>rd</sup> Ed.). Cambridge University Press, Great Britain, 331p.
- Brakstad, O.G., Aasbakk, K. and Maeland, J.A. 1992. Detection of *Staphylococcus aureus* by polymerase chain reaction amplification of the *nuc* gene. *J. Clin. Microbiol.* **30**: 1654-1660.
- Ciftci, A., Findik, A., Onuk, E.E. and Savasan, S. 2009. Detection of methicillin resistance and slime factor production of *Staphylococcus aureus* in bovine mastitis. *Brazilian J. Microbiol.* **40**: 254-261.
- Costa, A., Kay, I. and Palladino, S. 2004. Rapid detection of *mecA* and *nuc* genes in staphylococci by real-time multiplex polymerase chain reaction. *Diag. Microbiol. Infect. Dis.* **51**: 13-17.
- El-Razik, K.A.A., Abdelrahman, K.A., Ahmed, Y.F., Gomaa, A.M. and Eldebaky, H.A. 2010. Direct identification of major pathogens of the bubaline subclinical mastitis in Egypt using PCR. *J. Anim. Sci.* **6**: 652-660.
- Fadlelmula, A., Al Dughaym, A.M., Mohamed, G.E., Al Deib, M.K. and Al Zubaidy, A.J. 2009. Bovine mastitis: epidemiological, clinical and etiological study in a Saudi Arabian large dairy farm. *Bulg. J. Vet. Med.* **12**: 3.
- Fitzgerald, J.R., Hartigan, P.J., Meaney, W.J. and Smyth, C.J. 2000. Molecular population and virulence factor analysis of *Staphylococcus aureus* from bovine intramammary infection. *J. Appl. Microbiol.* **88**: 1028-1037.
- Fluit A.C. 2012. Livestock-associated *Staphylococcus aureus*. *Clin. Microbiol. Infect.* **18**: 735-44.
- Haveri, M., Roslof, A., Rantala, L. and Pyorala, S. 2007. Virulence genes of bovine *Staphylococcus aureus* from persistent and nonpersistent intramammary infections with different clinical characteristics. *J. Appl. Microbiol.* **103**: 993-1000.
- Jose, K.R., Vijayakumar, K., Justin, D.K., Shyma, V. H. and Ambily, R. 2021. Comparative efficacy of different treatment regimens against bovine mastitis caused by *Staphylococcus aureus*. *J. Vet. Anim. Sci.* **52**: 55-59.
- Kaneko, J. and Kamio, Y. 2004. Bacterial two-component and hetero-heptameric pore-forming cytolytic toxins: structures, pore-forming mechanism, and organization of the genes. *Biosci. Biotechnol. Biochem.* **68**: 981-1003.
- Lange, C.C., Brito, M.A., Reis, D.R., Machado, M.A., Guimaraes, A.S., Azevedo, A.L., Salles, E.B., Alvim, M.C., Silva, F.S. and Meurer, I.R. 2015. Species-level identification of staphylococci isolated from bovine mastitis in Brazil using partial 16S rRNA sequencing. *Vet. Microbiol.* **176**: 382-388.
- Miles, H., Lesser, W. and Sears P. 1992. The economic implications of bioengineered mastitis control. *J. Dairy Sci.* **75**: 596 - 605.
- Mitra, S.D., Velu, D., Bhuvana, M., Krithiga, N., Banerjee, A., Shome, R., Rahman, H., Ghosh, S.K. and Shome, B.R. 2013. *Staphylococcus aureus* spa type t267, clonal ancestor of bovine subclinical mastitis in India. *J. Appl. Microbiol.* **114**: 1604-1615.
- Nazir, N., Wani, S.A., Nyrah, Q., Farooq, S., Hassan, M.N. and Kashoo, Z.A. 2017. Virulence gene profile and antimicrobial

- resistance of *Staphylococcus aureus* isolated from bovine mastitis in Kashmir, India. *J. Appl. Nat. Sci.* **9**: 893-898.
- Pajic, M.J., Rasic, Z.B., Velebit, B.M., Bobos, S.F., Mihajlovic-Ukropina, M.M., Radinovic, M.Z., Galfi, A.L., Petkovic, J.M. and Trojancanec, S.I. 2014. The prevalence of methicillin resistance and Panton-Valentine Leukocidin synthesis genes in *Staphylococcus aureus* isolates of bovine and human origin. *Vet. Arhiv.* **84**: 205-14.
- Patel, K., Godden, S.M., Royster, E.E., Crooker, B.A., Johnson, T.J., Smith, E.A. and Sreevatsan, S. 2021. Prevalence, antibiotic resistance, virulence and genetic diversity of *Staphylococcus aureus* isolated from bulk tank milk samples of US dairy herds. *BMC genomics.* **22**: 1-13.
- Peton, V. and Le Loir, Y. 2014. *Staphylococcus aureus* in veterinary medicine. *Infect. Genet. Evol.* **21**: 602-615.
- Petti, C.A., Bosshard, P.P., Brandt, M.E., Clarridge, J.E., Feldblyum, T.V., Foxall, P., Furtado, M.R., Pace, N. and Procop, G. 2008. Interpretive criteria for identification of bacteria and fungi by DNA target sequencing; approved guideline. *Clinical and Laboratory Standards Institute (CLSI) Documents.* **28**: 19087-1898.
- Prashanth, K., Rao, K.R., Reddy, P.V., Saranathan, R. and Makki, A.R. 2011. Genotypic characterization of *Staphylococcus aureus* obtained from humans and bovine mastitis samples in India. *J. Glob. Inf. Dis.* **3**: 115.
- Prevost, G., Cribier, B., Couppie, P., Petiau, P., Supersac, G., Finck-Barbancon, V., Monteil, H., and Piemont, Y. 1995. Panton-Valentine leukocidin and gamma-hemolysin from *Staphylococcus aureus* ATCC 49775 are encoded by distinct genetic loci and have different biological activities. *Infect. Immun.* **63**: 4121– 4129.
- Quinn, P., Markey, B., Carter, M. and Carter, G.R. 2013. In: *Clinical Veterinary Microbiology*. (2nd Ed). Mosby, St. Louis, 514p.
- Rathish, R. L., Tresamol, P.V., Jacob, A., Nandu, T.G. and Saseendranath, M.R. 2015. Prevalence and antibiogram of *Pseudomonas* from bovine mastitis in Thrissur district. *J. Vet. Anim. Sci.* **46**: 81 – 83.
- Sasaki, T., Kikuchi, K., Tanaka, Y., Takahashi, N., Kamata, S. and Hiramatsu, K. 2007. Reclassification of phenotypically identified *Staphylococcus intermedius* strains. *J. Clin. Microbiol.* **45**: 2770-2778.
- Shah, H., Bhat, M.A., Nabi, B., Taku, A. and Badroo, G.A. 2020. Virulent gene characterization of methicillin sensitive *Staphylococcus aureus* (MSSA) and methicillin resistant *Staphylococcus aureus* (MRSA) from bovine mastitis. *Int. J. Curr. Microbiol. App. Sci.* **9**: 3471-3482.
- Shrivastava, N., Sharma, V., Shrivastav, A., Nayak, A. and Rai, A.K. 2018. Prevalence and characterization of Panton-Valentine leukocidin-positive *Staphylococcus aureus* in bovine milk in Jabalpur district of Madhya Pradesh, India. *Vet. World.* **11**: 16.
- Stephan, R., Annemuller, C., Hassan, A.A. and Lammler, C. 2001. Characterization of enterotoxigenic *Staphylococcus aureus* strains isolated from bovine mastitis in north-east Switzerland. *Vet. Microbiol.* **78**: 373-382.
- Verma, H., Rawat, S., Sharma, N., Jaiswal, V., Singh, R. and Harshit, V. 2018. Prevalence, bacterial etiology and antibiotic susceptibility pattern of bovine mastitis in Meerut. *J. Entomol. Zool. Stud.* **6**: 706-709.
- Vijayakumar, K. and Jose, K.R. 2021. Phenotypic and genotypic characterization of *Staphylococcus aureus* isolated from bovine mastitis for biofilm formation. *J. Anim. Res.* **11**: 249-255.
- Workineh, S., Bayleyegn, M., Mekonnen, H. and Potgieter, L.N.D. 2002. Prevalence and aetiology of mastitis in cows from two major Ethiopian dairies. *Trop. Anim. Health Prod.* **34**: 19-25.